



REVIEW

Open Access

Advances in the Pathogenesis of Alzheimer's Disease: Focusing on Tau-Mediated Neurodegeneration

Yale Duan¹, Suzhen Dong^{1,2}, Feng Gu¹, Yinghe Hu^{1,2} and Zheng Zhao^{1*}**Abstract**

In addition to senile plaques and cerebral amyloid angiopathy, the hyperphosphorylation of tau protein and formation of intraneuronal neurofibrillary tangles (NFTs) represents another neuropathological hallmark in AD brain. Tau is a microtubule-associated protein and localizes predominantly in the axons of neurons with the primary function in maintaining microtubules stability. When the balance between tau phosphorylation and dephosphorylation is changed in favor of the former, tau is hyperphosphorylated and the level of the free tau fractions elevated. The hyperphosphorylation of tau protein and formation of NFTs represent a characteristic neuropathological feature in AD brain. We have discussed the role of A β in AD in our previous review, this review focused on the recent advances in tau-mediated AD pathology, mainly including tau hyperphosphorylation, propagation of tau pathology and the relationship between tau and A β .

Keywords: Alzheimer's disease, Tau, A-beta, Tauopathy, Tau hyperphosphorylation, Intraneuronal neurofibrillary tangles

Review**Introduction**

In parallel with senile plaques and cerebral amyloid angiopathy, the hyperphosphorylation of tau protein and formation of intraneuronal neurofibrillary tangles (NFTs) represents another characteristic neuropathological feature in AD brain (see Figure 1). Tau is a microtubule-associated protein (MAP). Aberrantly phosphorylated tau is the main constituent of the aggregated paired helical filaments (PHF) that comprises NFT. Despite decades of intense research that strongly implicates NFT in underlying pathogenesis of AD and other neurodegenerative diseases (so called tauopathies) [1], there has been controversy as to whether NFTs or A β plaques are the primary cause of AD, and the interrelationship between these two pathologies remains largely elusive [2]. We have re-evaluated the role of A β in AD in our previous review [3]. In this review, we further discussed the

recent advances in tau-mediated AD pathology with focusing on the propagation of tau pathology, tau hyperphosphorylation and the relationship between tau and A β .

Physiological functions of tau

MAP tau localizes predominantly in the axons of neurons with the primary function in maintaining microtubules (MTs) stability. It is necessary for neurite outgrowth. Tau presents six main isoforms in the human brain (ranging between 352 and 441 amino acid residues), according to the alternative splicing, and differs by having 3 or 4 semi-conserved repeats of 31 residues in the MT-binding assembly domain and 0-2 insertions in the N-terminal projection domain [4,5]. Between the projection domain and the microtubule-binding domain lies a basic proline-rich region. MT-binding domain is important for promoting microtubule assembly, although it binds to microtubules only with low affinity. The various isoforms appear differentially during development. The ratio of 3R and 4R tau isoforms is 1:1 in most regions of the adult brain, and deviations from this ratio are characteristic of neurodegenerative tauopathies [6].

* Correspondence: zzhao@brain.ecnu.edu.cn

¹Key Laboratory of Brain Functional Genomics, Ministry of Education, Shanghai Key Laboratory of Brain Functional Genomics, East China Normal University, 3663 Zhongshan Road (N), Shanghai 200062, China
Full list of author information is available at the end of the article

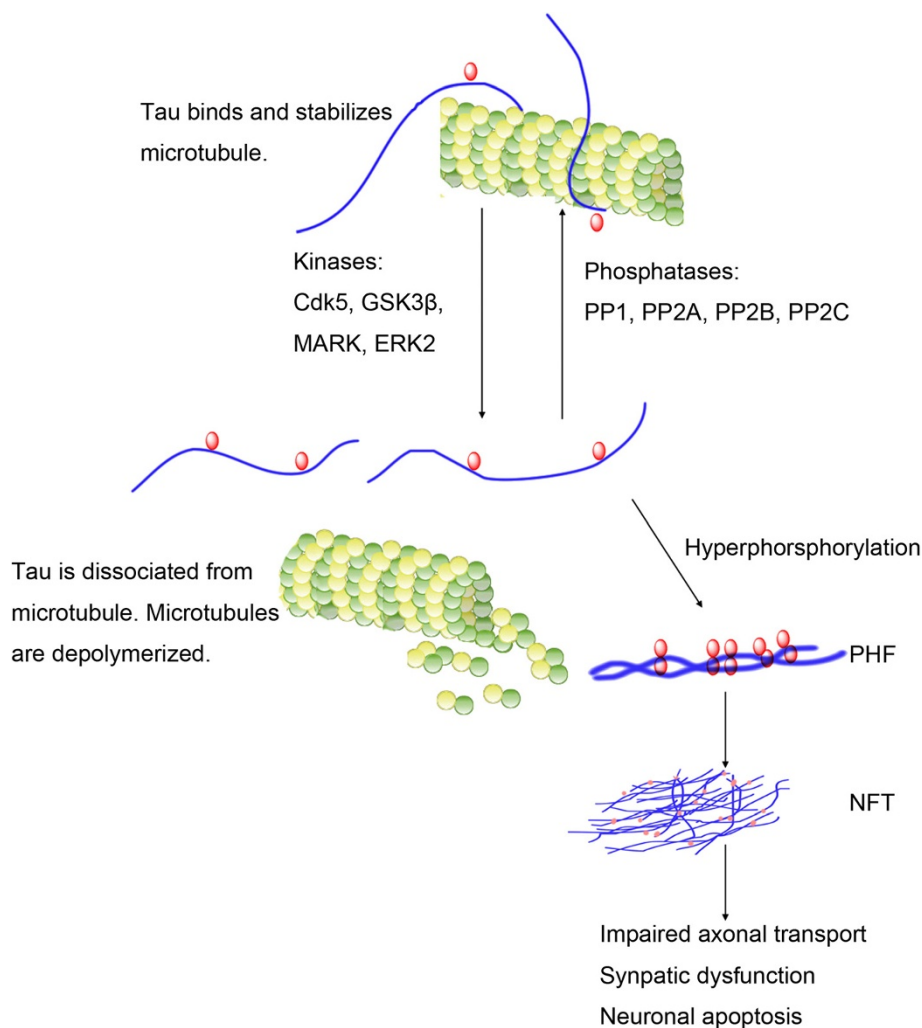


Figure 1 Tau-mediated neurodegeneration. Physiologically tau protein can bind and thereby stabilize microtubules (MTs). The attachment of tau to MT is regulated by its phosphorylation level. Phosphorylation of tau mediated by kinase (Cdk5, GSK3 β , MARK and ERK2) may lead to the detachment of tau from MT and hereby cause MT depolymerization. Conversely, phosphatase (PP1, PP2A, PP2B and PP2C) will reduce the phosphorylation level of tau and restore the binding ability of tau for MT. Such equilibrium between the roles of kinases and phosphatases is disrupted under pathological condition, and increase in the kinase activity and decrease in the phosphatase activity will cause tau hyperphosphorylation. Hyperphosphorylated tau protein is misfolded and forms β -sheet-containing structure paired helical filaments (PHFs). These structure transitions will lead to more organized aggregates, and eventually develop neurofibrillary tangles (NFT) inside neurons. NFT will impair normal axonal transport, disrupt synaptic plasticity, and finally induce cell loss.

The MT-binding ability of tau is post-translationally regulated primarily by serine/threonine-directed phosphorylation, which can effectively modulate the binding affinity of tau for MTs [7], although other post-translational modifications, such as glycosylation, may also have a direct impact on the dynamic equilibrium of tau on and off the MTs. Dynamic tau phosphorylation occurs during brain embryonic development [8]. It is substantially increased during the development of the fetal brain while decreased gradually in the adult brain [9]. Furthermore, tau has profound effects on axonal transport, function and viability of neurons and their

highly extended processes [10]. Tau is key to the sophisticated transport machinery that allows signaling molecules, trophic factors and other essential cellular constituents, including organelles (for example mitochondria and vesicles), to travel along the axons. Tau also interacts with various other proteins in addition to tubulin, including the SH3 domains of Src family tyrosine kinases [11,12]. These results strongly suggest that tau has a potential role in cell signaling. In addition, recent studies also demonstrated that tau plays functional roles in nucleolar organization and chromosome stability [13,14].

Pathological aggregation of tau

Free tau, PHF and NFTs

When the balance between tau phosphorylation and dephosphorylation is changed in favor of the former, tau is hyperphosphorylated and the level of the free tau fraction is elevated. Tau has been found to be phosphorylated at over 30 serine/threonine residues and approximately half of these are canonical sites for proline-directed protein kinases (PDPKs), suggesting important roles for PDPKs and protein phosphatases in the abnormal hyperphosphorylation of tau [15]. Hyperphosphorylated tau aggregates into NFTs through pretangles (nonfibrillary tau deposits) and PHF [16].

Phosphorylation and dephosphorylation of tau

Various other pathological events, including A β -mediated toxicity, as well as oxidative stress and inflammation, may be able to trigger or contribute (independently or in combination) to an abnormal detachment of tau from the MTs [17-20]. For example, it has been suggested that oxidative stress could be responsible for detrimental covalent modifications of tau, which include the formation of intermolecular disulphide bridges and tyrosine nitration. Such modifications are likely to cause misfolding, hyperphosphorylation and aggregation, and thereby contribute to abnormal disengagement of tau from MTs, as well as to the formation of aggregates. Although oxidative stress is often regarded as an upstream event relative to tau pathology, recent studies have revealed that pathological tau may interfere with mitochondrial function and induce oxidative stress [21-23]. In addition, lifetime stress, endoplasmic reticulum stress and hypersecreted glucocorticoids exposure also influence tau hyperphosphorylation [24,25].

Kinases of tau

Several lines of *in vitro* data have shown that many kinases are involved in phosphorylation of tau, though it is not yet clear if it is also physiologically or pathologically true *in vivo*. Nevertheless, cyclin-dependent kinase 5 (CDK5), glycogen synthase kinase 3 (GSK3), the microtubule-affinity-regulating kinase (MARK) and extracellular signal-regulated kinase 2 (ERK2) have received particular attention as potential targets for disease-modifying therapies using inhibitory compounds [7,26].

P35 and p39 proteins are expressed almost exclusively in postmitotic neurons and have been identified as CDK5 activators [27]. Elevated cellular calcium levels trigger the calpain-mediated cleavages of p35 and p39 to form the more stable p25 and p29 fragments [28,29]. Indeed, calpain activation, p25 accumulation and elevated CDK5 activity have all been observed directly in the AD brain [30,31]. This has also been evident in the

transgenic mice that overexpress human p25 that exhibit increased CDK5 activity, hyperphosphorylation of tau, neurofilament and cytoskeletal disturbances [32]. Inducible transgenic mice overexpressing p25 in the postnatal forebrain also exhibit neuronal loss and caspase-3 activation, accompanied by hyperphosphorylation of endogenous tau, accumulation of aggregated tau, and the progressively developed neurofibrillary pathology [33]. Together, these data suggest CDK5-p25 pathway is a crucial component of AD pathophysiology. Interestingly, mice overexpressing p35 as well as tau and CDK5 do not show increased tau phosphorylation, and the cdk5/p35 could not cause neurodegeneration in mouse brain, suggesting that cdk5/p35 might not be a major protein tau kinase [34].

Cdk5 modulates tau hyperphosphorylation via the inhibitory regulation of GSK3 [35]. GSK3 has two isoforms, GSK3 α and GSK3 β . In transfected mammalian cells, GSK3 α and GSK3 β could contribute to the formation of PHF [36,37]. Transgenic mice with elevated GSK3 β expressions show increased tau phosphorylation and deficits in spatial learning [38]. In newborn AD transgenic mouse models, knockdown of GSK3 α and GSK3 β reduces tau phosphorylation and tau misfolding, while the knockdown of GSK3 α , but not GSK3 β , leads to reduced senile plaques formation. These data demonstrate that GSK3 β only modulates NFT formation, while GSK3 α contributes to both senile plaques and NFT pathogenesis [39].

MARK phosphorylates tau on non-Ser/Thr-Pro sites and plays a crucial role in regulating tau's function. MARK selectively phosphorylates a KXGS motif, which is presented in each MT-binding domain of tau. Overexpression of MARK promotes tau phosphorylation at KXGS motifs and disrupts the microtubule array *in vivo* [40]. Although little is known so far about the upstream events that act through MARK to regulate tau phosphorylation, one recent study demonstrated that GSK3 β is substantially responsible for phosphorylating Ser-262 of tau through activation of MARK2 [41].

ERK2 is highly expressed in neurons and plays an important role in regulating tau functions and tau phosphorylation. This kinase can promote tau phosphorylation and thereby reduce the ability of tau in stabilizing microtubules [7].

Besides the roles either directly or indirectly in modulating tau phosphorylation, recent studies have revealed that the kinases mentioned above are also associated with APP cleavage. For instance, GSK3, especially GSK3 α , involves in APP processing, and the production of A β peptides can be significantly reduced by interfering APP cleavage at the gamma-secretase step with lithium, a GSK3 inhibitor [42]. Inhibition of GSK3 α may thus offer a new approach to reduce the formation of both amyloid

plaques and NFTs. CDK5-p25 can also modulate the production of A β by increasing APP phosphorylation at Thr668 [43].

Phosphatases of tau

It has been identified that a number of phosphatases, such as protein phosphatase (PP) 1, PP2A, PP2B and PP2C, could potentially drive the reverse and dephosphorylation of tau. Their activities were found to be decreased about 20-30% in AD brain [44].

PP2A is co-localized with tau and microtubules in the brain and is apparently the most active enzyme in dephosphorylation of tau. In AD brain, both the expression and activity of PP2A are decreased. Tau can be abnormally hyperphosphorylated if I1PP2A, a 249-amino acid long endogenous inhibitor of PP2A, is increased [45]. One recent study reported that PP2A could be inactivated via phosphorylation of its catalytic subunit at Y307. This PP2A inactivation can be mediated by A β deposition or estrogen deficiency in the AD brain. Moreover, the inactivation of PP2A consequently compromise dephosphorylation of abnormally hyperphosphorylated tau, therefore lead to neurofibrillary tangle formation [46].

Other post-translational modification of tau

In addition to tau phosphorylation, different types of post-translational modifications including acetylation, glycosylation, glycation, prolyl-isomerization, cleavage or truncation, nitration, polyamination, ubiquitination, sumoylation, oxidation and aggregation can regulation the function of tau [47-49]. Of these modification, tau acetylation is of great importance for tauopathy. Tau acetylation has recently been found to prevent p-tau from degradation and modulate the activities of kinase, implicating a central role in tauopathy [50].

The toxicity of tau

NFTs are considered to be responsible for the toxic effects of tau in AD for a long time, but recent findings suggest that this might not be all the fact. Santacruz et al. by using a strain of mice bearing a mutant human tau gene combined with regulatory sequences that allowed it to be turned off by the antibiotic doxycycline, demonstrated that the animals' memories are improved and the neuronal losses are halted when the mutant tau gene is switched off, but with no effect on NFT accumulation [51]. One study also showed that inhibition of tau phosphorylation is able to prevent the typical motor deficits and markedly reduce soluble aggregated hyperphosphorylated tau in the tau transgenic mice [52], suggesting that PHF or other soluble lower-mass hyperphosphorylated tau aggregates are neurotoxic. Meanwhile, increasing evidence has revealed that tau-mediated neurodegeneration may result from the

combination of gain-of-toxic function acquired by the aggregates or their precursors and the detrimental effects that arise from the loss of the normal function(s) of tau in the disease state [53,54].

Propagation of tau pathology

NFTs have a hierarchical pattern of accumulation in vulnerable neurons. The neurons in layer II of the entorhinal cortex (EC-II) are considered as the first to be affected. Later, the lesions appear to spread to limbic and association cortices [55]. However, the exact mechanism involved in the pattern of propagation is incompletely understood. Recently, several studies have showed that the intracellular protein aggregates of tau can spread by a prion-like mechanism in the brain. The extracellular tau aggregates can enter cells through endocytosis and trigger the misfolding and aggregation of intracellular tau in cell culture experiments [56-58]. In another study [59], de Calignon et al. generated a transgenic mouse model in which overexpression of human mutant tau (P301L) is restricted to EC-II (named rTgTauEC mouse) to investigate the disease progression. They found that tau pathology progresses from the EC neurons expressing the human transgene to the nearby neurons, and then to neurons located downstream in the synaptic circuit, for instance the dentate gyrus, hippocampus, and cingulate cortex [59]. These findings provide useful information for understanding the hierarchical patterns of tau-mediated neurodegeneration in AD.

Tau and A β

Both senile plaques and NFTs are predominant pathologic characteristics of AD. Connections between A β toxicity and tau pathology have repeatedly been proposed. However, the underlying mechanisms have not yet been fully established, and this remains one of the most challenging conundrums of AD research.

It is increasingly recognized that reduction in tau levels can alleviate memory loss in the AD mouse model. Mucke and his colleagues found that decreasing endogenous tau prevents behavioral deficits in transgenic mice with mutant APP, without altering their high A β levels. Tau reduction also protected both APP transgenic AD model and nontransgenic mice against excitotoxicity. The absence of tau somehow prevents the behavioral deficits that would otherwise occur in animals [60]. New lines of investigation support the notion that tau malfunction, in addition to being independently capable of producing neurodegeneration even in the absence of A β deposits or other pathological events, could be a key mediator of neurodegeneration in response to other upstream events, including A β -induced neurotoxicity [20]. It has been shown that A β can bind to tau and form a stable complex both in vitro and in AD brain [61]. The

complex enhances tau phosphorylation via GSK-3 β signaling, suggesting that A β lies in the upstream of tau pathology. This has been further supported by the study of Bolmont et al. in which intracerebral injection of brain extracts from APP transgenic mice induced the formation of NFT in mutant tau transgenic mice [62]. The mutant tau and APP double transgenic mice exhibit more significant increasing in tau pathology than the mutant tau transgenic mice predominantly in the area with high amyloid burden, while the double transgene do not lead to up-regulation of amyloid load as compared with the mutant APP transgenic mice. A β and phosphorylated-tau have been observed to be co-localized in synaptic terminals of AD brains [63]. A β can result in the transcriptional up-regulation of a gene named dual-specificity tyrosine-regulated kinase 1A (DYRK1A), further leading to tau phosphorylation [64]. Moreover, microtubule disassembly, one of pathological functions of tau, is initiated by prefibrillar A β [18]. Also, tau is required for the cytotoxicity of hybrid oligomers formed by A β _{3(pE)-42} and A β ₁₋₄₂ [65]. Recent studies showed that tau deficiency in *tau*^{-/-} mice and truncated tau in transgenic mice both lead to disruption of postsynaptic targeting of Fyn kinase and attenuation of A β toxicity, indicating that tau is a mediator of A β toxicity [66].

All these suggest that A β may drive tau pathology, and tau can mediate A β toxicity, implicating a cooperation between A β and tau for AD pathology. In this regard, it has been demonstrated that A β and tau could synergistically impair mitochondrial respiration in a triple transgenic Alzheimer's disease mice [67,68]. However, the exact mechanisms underlying such an interaction of A β and tau need further investigation.

Conclusion

As a most common neurodegenerative disorder, AD characterized by hyperphosphorylation of tau protein and formation of NFTs are collectively termed "tauopathy". This review highlights the recent advances in tau-mediated AD pathology, including tau hyperphosphorylation, propagation of tau pathology and the relationship between tau and A β . Tau plays an unequivocal role in AD, but the mechanisms of tau that induce dysfunction and death of neurons remain incompletely understood. Future researches can focus on the precise mechanisms of tau involved in the disease pathogenesis, which may eventually lead to the development of new therapeutic strategies for tauopathies of AD.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

YLD, SZD and FG collected the reference materials and drafted the manuscript. YHH and ZZ conceived of the study, and participated in its

design and coordination and helped to draft the manuscript. All authors read and approved the final manuscript.

Acknowledgements

This work was supported by the grants from the National Natural Science Foundation of China (No. 31171019, No. 81173108, No. 31000574 and No. 31200820), and the Opening Projects of Shanghai Key Laboratory of Brain Functional Genomics and Key Laboratory of Brain Functional Genomics (East China Normal University), Ministry of Education.

Author details

¹Key Laboratory of Brain Functional Genomics, Ministry of Education, Shanghai Key Laboratory of Brain Functional Genomics, East China Normal University, 3663 Zhongshan Road (N), Shanghai 200062, China. ²Shanghai Engineering Research Center for Molecular Therapeutics and New Drug Development, East China Normal University, Shanghai 200062, China.

Received: 1 November 2012 Accepted: 11 December 2012

Published: 15 December 2012

References

1. Lee VM, Goedert M, Trojanowski JQ: Neurodegenerative tauopathies. *Annu Rev Neurosci* 2001, **24**:1121-1159.
2. Lee VMY: Tauists and baptists United-Well Almost! *Science* 2001, **293**:1446-1447.
3. Dong SZ, Duan YL, Gu F, Hu YH, Zhao Z: Advances in the pathogenesis of Alzheimer's disease: a re-evaluation of amyloid cascade hypothesis. *Translational Neurodegeneration* 2012, **1**:18.
4. Goedert M, Spillantini MG, Jakes R, Rutherford D, Crowther RA: Multiple isoforms of human microtubule-associated protein tau: sequences and localization in neurofibrillary tangles of Alzheimer's disease. *Neuron* 1989, **3**:519-526.
5. Lee G, Cowan N, Kirschner M: The primary structure and heterogeneity of tau protein from mouse brain. *Science* 1988, **239**:285-288.
6. Hong M, Zhukareva V, Vogelsberg-Ragaglia V, Wszolek Z, Reed L, Miller BL, Geschwind DH, Bird TD, McKeel D, Goate A, et al: Mutation-specific functional impairments in distinct tau isoforms of hereditary FTDP-17. *Science* 1998, **282**:1914-1917.
7. Mazanetz MP, Fischer PM: Untangling tau hyperphosphorylation in drug design for neurodegenerative diseases. *Nat Rev Drug Discov* 2007, **6**:464-479.
8. Brion JP, Octave JN, Couck AM: Distribution of the phosphorylated microtubule-associated protein tau in developing cortical neurons. *Neuroscience* 1994, **63**:895-909.
9. Yu Y, Run X, Liang Z, Li Y, Liu F, Liu Y, Iqbal K, Grundke-Iqbal I, Gong CX: Developmental regulation of tau phosphorylation, tau kinases, and tau phosphatases. *J Neurochem* 2009, **108**:1480-1494.
10. Wang JZ, Liu F: Microtubule-associated protein tau in development, degeneration and protection of neurons. *Prog Neurobiol* 2008, **85**:148-175.
11. Lee G, Newman ST, Gard DL, Band H, Panchamoorthy G: Tau interacts with src-family non-receptor tyrosine kinases. *J Cell Sci* 1998, **111**(Pt 21):3167-3177.
12. Reynolds CH, Garwood CJ, Wray S, Price C, Kellie S, Perera T, Zvelebil M, Yang A, Sheppard PW, Varndell IM, et al: Phosphorylation regulates tau interactions with SH3 domains of phosphatidylinositol-3-kinase, phospholipase cgamma 1, GRB2 and SRC-family kinases. *J Biol Chem* 2008, **283**(26):18177-86.
13. Sjoberg MK, Shestakova E, Mansuroglu Z, Maccioni RB, Bonnefoy E: Tau protein binds to pericentromeric DNA: a putative role for nuclear tau in nucleolar organization. *J Cell Sci* 2006, **119**:2025-2034.
14. Rossi G, Dalpra L, Crosti F, Lissoni S, Sciacca FL, Catania M, Di Fede G, Mangieri M, Giaccone G, Croci D, Tagliavini F: A new function of microtubule-associated protein tau: involvement in chromosome stability. *Cell Cycle* 2008, **7**:1788-1794.
15. Wang JZ, Grundke-Iqbal I, Iqbal K: Kinases and phosphatases and tau sites involved in Alzheimer neurofibrillary degeneration. *Eur J Neurosci* 2007, **25**:59-68.
16. Congdon EE, Kim S, Bonchak J, Songrug T, Matzavinos A, Kuret J: Nucleation-dependent Tau Filament Formation: the importance of dimerization and an estimation of elementary rate constants. *J Biol Chem* 2008, **283**:13806-13816.

17. Andersen JK: **Oxidative stress in neurodegeneration: cause or consequence?** *Nat Med* 2004, **10**(Suppl):S18–25.
18. King ME, Kan HM, Baas PW, Erisir A, Glabe CG, Bloom GS: **Tau-dependent microtubule disassembly initiated by prefibrillar beta-amyloid.** *J Cell Biol* 2006, **175**:541–546.
19. Moreira PI, Smith MA, Zhu X, Nunomura A, Castellani RJ, Perry G: **Oxidative stress and neurodegeneration.** *Ann N Y Acad Sci* 2005, **1043**:545–552.
20. Rapoport M, Dawson HN, Binder LI, Vitek MP, Ferreira A: **Tau is essential to beta-amyloid-induced neurotoxicity.** *PNAS* 2002, **99**:6364–6369.
21. David DC, Hauptmann S, Scherping I, Schuessel K, Keil U, Rizzo P, Ravid R, Drose S, Brandt U, Muller WE, et al: **Proteomic and functional analyses reveal a mitochondrial dysfunction in P301L tau transgenic mice.** *J Biol Chem* 2005, **280**:23802–23814.
22. Revett TJ, Baker GB, Jhamandas J, Kar S: **Glutamate system, amyloid ss peptides and tau protein: functional interrelationships and relevance to Alzheimer disease pathology.** *J Psychiatry Neurosci* 2012, **37**:110190.
23. DuBoff B, Götz J, Feany Mel B: **Tau Promotes Neurodegeneration via DRP1 Mislocalization In Vivo.** *Neuron* 2012, **75**:618–632.
24. Sotiropoulos I, Catania C, Pinto LG, Silva R, Pollerberg GE, Takashima A, Sousa N, Almeida OF: **Stress acts cumulatively to precipitate Alzheimer's disease-like tau pathology and cognitive deficits.** *J Neurosci* 2011, **31**:7840–7847.
25. Ho YS, Yang X, Lau JC, Hung CH, Wuwongse S, Zhang Q, Wang J, Baum L, So KF, Chang RC: **Endoplasmic reticulum stress induces tau pathology and forms a vicious cycle: implication in Alzheimer's disease pathogenesis.** *J Alzheimers Dis* 2012, **28**:839–854.
26. Kim I, Park EJ, Seo J, Ko SJ, Lee J, Kim CH: **Zinc stimulates tau S214 phosphorylation by the activation of Raf/mitogen-activated protein kinase-kinase/extracellular signal-regulated kinase pathway.** *Neuroreport* 2011, **22**:839–844.
27. Tsai LH, Delalle I, Caviness VS Jr, Chae T, Harlow E: **p35 is a neural-specific regulatory subunit of cyclin-dependent kinase 5.** *Nature* 1994, **371**:419–423.
28. Hashiguchi M, Saito T, Hisanaga S, Hashiguchi T: **Truncation of CDK5 activator p35 induces intensive phosphorylation of Ser202/Thr205 of human tau.** *J Biol Chem* 2002, **277**:44525–44530.
29. Patzke H, Tsai LH: **Calpain-mediated cleavage of the cyclin-dependent kinase-5 activator p39 to p29.** *J Biol Chem* 2002, **277**:8054–8060.
30. Lee KY, Clark AW, Rosales JL, Chapman K, Fung T, Johnston RN: **Elevated neuronal Cdc2-like kinase activity in the Alzheimer disease brain.** *Neurosci Res* 1999, **34**:21–29.
31. Tseng HC, Zhou Y, Shen Y, Tsai LH: **A survey of Cdk5 activator p35 and p25 levels in Alzheimer's disease brains.** *FEBS Lett* 2002, **523**:58–62.
32. Ahljanian MK, Barrezuela NX, Williams RD, Jakowski A, Kowicz KP, McCarthy S, Coskran T, Carlo A, Seymour PA, Burkhardt JE, et al: **Hyperphosphorylated tau and neurofilament and cytoskeletal disruptions in mice overexpressing human p25, an activator of cdk5.** *PNAS* 2000, **97**:2910–2915.
33. Cruz JC, Tseng HC, Goldman JA, Shih H, Tsai LH: **Aberrant Cdk5 activation by p25 triggers pathological events leading to neurodegeneration and neurofibrillary tangles.** *Neuron* 2003, **40**:471–483.
34. Van den Haute C, Spittaels K, Van Dorpe J, Lasrado R, Vandezande K, Laenen I, Geerts H, Van Leuven F: **Coexpression of human cdk5 and its activator p35 with human protein tau in neurons in brain of triple transgenic mice.** *Neurobiol Dis* 2001, **8**:32–44.
35. Plattner F, Angelo M, Giese KP: **The roles of cyclin-dependent kinase 5 and glycogen synthase kinase 3 in tau hyperphosphorylation.** *J Biol Chem* 2006, **281**:25457–25465.
36. Lovestone S, Reynolds CH, Latimer D, Davis DR, Anderton BH, Gallo JM, Hanger D, Mulot S, Marquardt B, Stabel S, et al: **Alzheimer's disease-like phosphorylation of the microtubule-associated protein tau by glycogen synthase kinase-3 in transfected mammalian cells.** *Curr Biol* 1994, **4**:1077–1086.
37. Sperber BR, Leight S, Goedert M, Lee VM: **Glycogen synthase kinase-3 beta phosphorylates tau protein at multiple sites in intact cells.** *Neurosci Lett* 1995, **197**:149–153.
38. Hernandez F, Borrell J, Guaza C, Avila J, Lucas JJ: **Spatial learning deficit in transgenic mice that conditionally over-express GSK-3beta in the brain but do not form tau filaments.** *J Neurochem* 2002, **83**:1529–1533.
39. Hurtado DE, Molina-Porcel L, Carroll JC, Macdonald C, Aboagye AK, Trojanowski JQ, Lee VM: **Selectively silencing GSK-3 isoforms reduces plaques and tangles in mouse models of Alzheimer's disease.** *J Neurosci* 2012, **32**:7392–7402.
40. Drewes G, Ebner A, Preuss U, Mandelkow EM, Mandelkow E: **MARK, a novel family of protein kinases that phosphorylate microtubule-associated proteins and trigger microtubule disruption.** *Cell* 1997, **89**:297–308.
41. Kosuga S, Tashiro E, Kajioaka T, Ueki M, Shimizu Y, Imoto M: **GSK-3beta directly phosphorylates and activates MARK2/PAR-1.** *J Biol Chem* 2005, **280**:42715–42722.
42. Phiel CJ, Wilson CA, Lee VM, Klein PS: **GSK-3alpha regulates production of Alzheimer's disease amyloid-beta peptides.** *Nature* 2003, **423**:435–439.
43. Lee MS, Kao SC, Lemere CA, Xia W, Tseng HC, Zhou Y, Neve R, Ahljanian MK, Tsai LH: **APP processing is regulated by cytoplasmic phosphorylation.** *J Cell Biol* 2003, **163**:83–95.
44. Gong CX, Shaikh S, Wang JZ, Zaidi T, Grundke-Iqbal I, Iqbal K: **Phosphatase activity toward abnormally phosphorylated tau: decrease in Alzheimer disease brain.** *J Neurochem* 1995, **65**:732–738.
45. Chen S, Li B, Grundke-Iqbal I, Iqbal K: **11PP2A affects tau phosphorylation via association with the catalytic subunit of protein phosphatase 2A.** *J Biol Chem* 2008, **283**:10513–10521.
46. Liu R, Zhou XW, Tanila H, Bjorkdahl C, Wang JZ, Guan ZZ, Cao Y, Gustafsson JA, Winblad B, Pei JJ: **Phosphorylated PP2A (tyrosine 307) is associated with Alzheimer neurofibrillary pathology.** *J Cell Mol Med* 2008, **12**:241–257.
47. Martin L, Latypova X, Terro F: **Post-translational modifications of tau protein: Implications for Alzheimer's disease.** *Neurochem Int* 2011, **58**:458–471.
48. Wang JZ, Xia YY, Grundke-Iqbal I, Iqbal K: **Abnormal Hyperphosphorylation of Tau: Sites, Regulation, and Molecular Mechanism of Neurofibrillary Degeneration.** *J Alzheimers Dis* 2012, doi:10.3233/JAD-2012-129031.
49. Irwin DJ, Cohen TJ, Grossman M, Arnold SE, Xie SX, Lee VMY, Trojanowski JQ: **Acetylated tau, a novel pathological signature in Alzheimer's disease and other tauopathies.** *Brain* 2012, **135**:807–818.
50. Min SW, Cho SH, Zhou Y, Schroeder S, Haroutunian V, Seeley WW, Huang EJ, Shen Y, Masliah E, Mukherjee C, et al: **Acetylation of tau inhibits its degradation and contributes to tauopathy.** *Neuron* 2010, **67**:953–966.
51. Santacruz K, Lewis J, Spire T, Paulson J, Kotilinek L, Ingelsson M, Guimaraes A, DeTure M, Ramsden M, McGowan E, et al: **Tau suppression in a neurodegenerative mouse model improves memory function.** *Science* 2005, **309**:476–481.
52. Le Corre S, Klafki HW, Plesnila N, Hubinger G, Obermeier A, Sahagun H, Monse B, Seneci P, Lewis J, Eriksen J, et al: **An inhibitor of tau hyperphosphorylation prevents severe motor impairments in tau transgenic mice.** *PNAS* 2006, **103**:9673–9678.
53. Trojanowski JQ, Lee VM: **Pathological tau: a loss of normal function or a gain in toxicity?** *Nat Neurosci* 2005, **8**:1136–1137.
54. Winklhofer KF, Tatzelt J, Haass C: **The two faces of protein misfolding: gain- and loss-of-function in neurodegenerative diseases.** *EMBO J* 2008, **27**:336–349.
55. Hyman BT, Trojanowski JQ: **Consensus recommendations for the postmortem diagnosis of Alzheimer disease from the National Institute on Aging and the Reagan Institute Working Group on diagnostic criteria for the neuropathological assessment of Alzheimer disease.** *J Neuropathol Exp Neurol* 1997, **56**:1095–1097.
56. Frost B, Jacks RL, Diamond MI: **Propagation of tau misfolding from the outside to the inside of a cell.** *J Biol Chem* 2009, **284**:12845–12852.
57. Guo JL, Lee VM: **Seeding of normal Tau by pathological Tau conformers drives pathogenesis of Alzheimer-like tangles.** *J Biol Chem* 2011, **286**:15317–15331.
58. Nonaka T, Watanabe ST, Iwatsubo T, Hasegawa M: **Seeded aggregation and toxicity of [alpha]-synuclein and tau: cellular models of neurodegenerative diseases.** *J Biol Chem* 2010, **285**:34885–34898.
59. de Calignon A, Polydoro M, Suarez-Calvet M, William C, Adamowicz DH, Kopeikina KJ, Pitstick R, Sahara N, Ashe KH, Carlson GA, et al: **Propagation of tau pathology in a model of early Alzheimer's disease.** *Neuron* 2012, **73**:685–697.
60. Roberson ED, Scarce-Levie K, Palop JJ, Yan F, Cheng IH, Wu T, Gerstein H, Yu GQ, Mucke L: **Reducing endogenous tau ameliorates amyloid beta-induced deficits in an Alzheimer's disease mouse model.** *Science* 2007, **316**:750–754.
61. Guo JP, Arai T, Miklosy J, McGeer PL: **Abeta and tau form soluble complexes that may promote self aggregation of both into the insoluble forms observed in Alzheimer's disease.** *PNAS* 2006, **103**:1953–1958.

62. Bolmont T, Clavaguera F, Meyer-Luehmann M, Herzig MC, Radde R, Staufenbiel M, Lewis J, Hutton M, Tolnay M, Jucker M: **Induction of tau pathology by intracerebral infusion of amyloid-beta -containing brain extract and by amyloid-beta deposition in APP x Tau transgenic mice.** *Am J Pathol* 2007, **171**:2012–2020.
63. Fein JA, Sokolow S, Miller CA, Vinters HV, Yang F, Cole GM, Gyllys KH: **Co-localization of amyloid beta and tau pathology in Alzheimer's disease synaptosomes.** *Am J Pathol* 2008, **172**:1683–1692.
64. Kimura R, Kamino K, Yamamoto M, Nuripa A, Kida T, Kazui H, Hashimoto R, Tanaka T, Kudo T, Yamagata H, *et al*: **The DYRK1A gene, encoded in chromosome 21 Down syndrome critical region, bridges between beta-amyloid production and tau phosphorylation in Alzheimer disease.** *Hum Mol Genet* 2007, **16**:15–23.
65. Nussbaum JM, Schilling S, Cynis H, Silva A, Swanson E, Wangsanut T, Tayler K, Wiltgen B, Hatami A, Ronicke R, *et al*: **Prion-like behaviour and tau-dependent cytotoxicity of pyroglutamylated amyloid-beta.** *Nature* 2012, **485**:651–655.
66. Ittner LM, Ke YD, Delerue F, Bi M, Gladbach A, van Eersel J, Wolfing H, Chieng BC, Christie MJ, Napier IA, *et al*: **Dendritic function of tau mediates amyloid-beta toxicity in Alzheimer's disease mouse models.** *Cell* 2010, **142**:387–397.
67. Rhein V, Song X, Wiesner A, Ittner LM, Baysang G, Meier F, Ozmen L, Bluethmann H, Drose S, Brandt U, *et al*: **Amyloid-beta and tau synergistically impair the oxidative phosphorylation system in triple transgenic Alzheimer's disease mice.** *PNAS* 2009, **106**:20057–20062.
68. Ittner LM, Götz J: **Amyloid- β and tau — a toxic pas de deux in Alzheimer's disease.** *Nat Rev Neurosci* 2010, **12**:65–72.

doi:10.1186/2047-9158-1-24

Cite this article as: Duan *et al.*: Advances in the Pathogenesis of Alzheimer's Disease: Focusing on Tau-Mediated Neurodegeneration. *Translational Neurodegeneration* 2012 **1**:24.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

